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Complete genome sequence and comparative analysis of two potential probiotics *Bacillus subtilis* isolated from honey and honeybee microbiomes

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Abstract

Background: We have previously isolated *Bacillus subtilis* HMNig-2 and MENO2 strains, from honey and the honeybee gut microbiome respectively, and demonstrated these strains to produce levansucrase with potential probiotics characteristics. Here we report their complete genome sequence and comparative analysis with other and other *B. subtilis* strains.

Results: The complete genome sequences of *Bacillus subtilis* HMNig-2 and MENO2 were de novo assembled from MiSeq paired-end sequence reads and annotated using the RAST tool. Whole-genome alignments were performed to identify functional differences associated with their potential use as probiotics.

Conclusions: The comparative analysis and the availability of the genome sequence of these two strains will provide comprehensive analysis about the diversity of these valuable *Bacillus* strains and the possible impact of the environment on bacterial evolution.

Significance and impact of study: We introduce complete genome sequence of two new *B. subtilis* strains HMNig-2 and MENO2 with probiotic potential and as cell factories for the production of levan and other valuable components for pharmaceutical and industrial applications

Keywords: Genome sequence, *Bacillus subtilis*, Probiotics, Honey, Honeybee

Background

Honey has been used as a preserve and to treat infection for four millennia [1]. Honey is known for its antibacterial activities, which relate to the properties of viscosity, acidity, and the presence of inimical substances such as H₂O₂, flavonoids, and phenolic acids [2]. However, it has

been documented that natural honey also contains a small number of microorganisms that constitute the indigenous microflora and which have probiotic potential [3]. Thus, we have used honey and the honey bee as sources of levansucrase producing probiotic bacteria. Two candidate probiotic bacteria that produce levansucrase were isolated, HMNig-2 and MENO2, which were identified as *Bacillus subtilis* species [4, 5].

The natural characteristics of honey that include pH, moisture content, antimicrobial components and sugar concentration mark it as a suitable environment for the persistence of spore-forming bacteria. Even though soil is considered the principal habitat for *Bacillus* species,

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they have been detected in honey [6] and the feces of bee larvae [7, 8].

Bacillus strains have been selected for use in probiotic supplements for human and animal consumption [9–12]. As these bacteria form spores, they present advantages over other common probiotic species with the ability to tolerate high temperatures and sustain viability for long periods without the need for cooling or freezing [12, 13]. *Bacillus* species are also widely used in industrial biotechnology as cell factories to produce a multiplicity of metabolites including antibiotics, bioinsecticides, enzymes, and lipopeptides [14–17].

Levansucrase is an extracellular enzyme encoded by the *sacB* gene, which belongs to the glycoside hydrolase (GH) 68 family produced by several Gram-positive and Gram-negative microbial species. In the presence of high sucrose concentrations, *B. subtilis* produces levan by means levansucrase activity. Industrial applications of levan have been proposed for food and pharmaceutical use. In food applications, levan has been used as an emulsifier, formulation aid, stabilizer and thickener, surface-finishing agent, encapsulating agent, and a carrier for flavor and fragrances [18]. In medicine, levan has shown promise as a plasma substitute, drug activity prolongator, and antihyperlipidemic agent [19]. Although a number of microorganisms are capable of producing levan, including *Bacillus polymyxa* [20], *Acetobacter xylinum* [21], *Lactobacillus sanfranciscensis* [22], *Leuconostoc mesenteroides* [23], *Microbacterium laevaniformans* [24], *Zymomonas mobilis* [25], *B. subtilis* [26], *B. amyloliquefaciens* [27], and *Pseudomonas syringae* [28]; relatively few have been demonstrated to produce levan suitable for food use or in yields of necessary to support commercial production. These restrictions have limited the application of levan.

Here we announce the complete genome sequence and the comparative analysis of the levansucrase producing *B. subtilis* strains HMNig-2 and MENO2 which have been isolated from honey and the honey bee microbiomes. The comparative analysis of these two strains and the availability of their genome sequences will provide a basis for their application as probiotics as well as cell factories for the levan production and other valuable components.

Methods

Strains and growth conditions

B. subtilis strains HNMig-2 and MENO2 are isolated previously in our laboratory from honey samples and honey bee microbiome [4]. *B. subtilis* strains were grown at 37 °C in Luria Bertani (LB) broth or agar plates made by adding 1.5% agar to the LB medium.

Genomic DNA extraction

The bacterial cultures were subjected to genomic DNA extraction, using AxyPrep bacterial genomic DNA mini-prep kit (Axygen Biosciences, Union City, CA, USA) according to the manufacturer's instructions and the quality checked using a NanoDrop instrument (ThermoFisher, Loughborough, UK) and by 1% agarose gel electrophoresis.

Genome sequencing

Genome DNA from libraries prepared using the Illumina Nextera™ tagmentation protocol and run on the MiSeq using an Illumina v3 cassette (Illumina, USA). The genome sequences were de novo assembled from approximately 3.2 million 80 to 250-bp paired-end reads each for HMNig-2 and MENO2 using CLC Genomics Workbench v10.01 and v12.03 (Qiagen Bioinformatics, Denmark). DNA sequence contigs were aligned to the rhizosphere sourced *B. subtilis* strains GQJK2 (NCBI accession CP020367) and TLO3 (NCBI accession CP021169) using whole-genome alignment within CLC Genomics Workbench and used in support of genome sequence assembly.

Genome annotation and comparison

The complete genomes of *B. subtilis* strains HMNig-2 and MENO2 were annotated using the RAST server [29]. The CGView Comparison Tool was used to generate the maps [30] using BLAST comparisons with an Expect value of 0.1. The first and second rings show the locations of protein coding, tRNA, and rRNA genes on the forward and reverse strands, respectively. The black plot depicts GC content with the peaks extending towards the outside of the circle representing GC content above the genome average, whereas those extending towards the center mark segments with GC content lower than the genome average. The innermost plot depicts GC skew, and both base composition plots were generated using a sliding window of 50,000 nt. The complete genome sequences of the two *Bacillus subtilis* strains have been deposited in the GenBank database.

Phylogenetic and comparative analysis of levansucrase production

The nucleotide and amino acid sequences encoded by the levansucrase operon from the *B. subtilis* isolates were analyzed by using the BLAST server against the Swiss-Prot protein database [<http://www.expasy.org/tools/blast/>], and the sequences aligned with the CLUSTALW interface in MEGA7.0 (<http://www.megasoftware.net/>; pairwise alignment gap opening penalty, 10; gap extension penalty, 0.1; multiple alignment gap opening penalty, 10; gap extension penalty, 0.2). A structure-based alignment was performed with MUSCLE, and the resulting alignment was further refined manually.

Results

General genome features

The results of the genome sequence of the two strains indicate similarities but they differ in size and coding capacity. Their general genome features are presented in Table 1, which show that the genomes of *B. subtilis* HMNig-2 and MENO2 contain 4,188,942 and 4,084,669 bases predicting 4239 and 4127 CDSs respectively, and with average of GC contents of 43.6 and 43.8% respectively.

Comparison with database genome sequences of *Bacillus* species revealed that they exhibit 85–95% DNA sequence identity between the CDSs present in these strains and many *B. subtilis* strains. The complete genome sequences for the two strains are available in GenBank and assigned the nucleotide accession numbers CP031784 for HMNig-2 and CP031783 for MENO2.

Comparative study of the *B. subtilis* HMNig-2 and *B. subtilis* MENO2 genomes

The complete genomes of HMNig-2 and MENO2 were analyzed by genome Blast to find the closet neighbors for each strain. From these results, phylogenetic trees were generated which are presented in Fig. 1. The complete genomes of HMNig-2 and MENO2 were compared to 100 of the closest *B. subtilis* strains available to detect regions of similarity among them. These results are shown in Fig. 2a and b.

Levansucrase gene

The nucleotide sequences of the levansucrase genes encoded by *sacB* were identified in the genome sequences of HMNig-2 and MENO2 as ORFs of 1419 bp and 1417 bp that code for proteins of 473 and 472 amino acids respectively. The nucleotide sequences of the *sacB* genes were aligned with those of *B. subtilis* isolates from different sources (soil, food, gut, sea, plant) using clustal Omega, and from which a phylogenetic tree was generated (Fig. 3). The phylogenetic tree is formed of five clades, although the sources of isolation do not coincide with the clade distributions, and notably HMNig-2 and MENO2 are not located in the same clade. BLAST searches at NCBI for these sequences at the nucleotide and amino acid sequence level showed

high sequence identity with *sacB* from Gram-positive microorganisms such as *B. subtilis* (99%) (strains: ge28, VV2, YP1, TLO3, and BS16045), *B. licheniformis* strain SRCM101441 (98%), *B. amyloliquefaciens* (83%) (strains: LM2303, L-S60, L-H15, LFB112, and Y14), and *Lactobacillus reuteri* (41%) (accession numbers: ABQ01720.1 and AVK93185.1). The sequences presented show reduced identity to *sacB* sequences from Gram-negative microorganisms including *Zymomonas mobilis* strains (26%) (WP_013934123.1, WP_011240294, AAG29870.1, AAV88999.2) and *Erwinia amylovora* (26%). The amino acid sequences of *sacB* gene from *B. subtilis* HMNig-2 and MENO2 showed high similarities (99%) to glycoside hydrolase family 68 protein of *B. subtilis* and multispecies strains (accession numbers: WP_069703641.1, WP_086344351.1, WP_032726907.1, WP_087987552.1, WP_015251171.1, WP_014115253.1, WP_024571701.1). High sequence identity was also noted for the amino acid sequences of *B. subtilis* levansucrase for which crystallographic structures are available (PDB accession numbers: 1OYG_A, 3BYJ_A, 3BYK_A) and *B. subtilis* levansucrase of biotechnological use (accession numbers: KIU05730.1, KFI02859.1, AAN75494.1, CBI68350.1).

The levansucrase sequences of both HMNig-2 and MENO2 exhibit the key features of the *Bacillus* enzymes: a signal peptide MNIKKFAKQATVLTFTTALLAGGATQAFA corresponding to residues 1-29, 3 active sites residues corresponding with D86, D247, and E342. In addition, they have a conserved residue at D339, which is known to bind Ca^{2+} in *B. subtilis* levansucrase. The levansucrase reading frame of HMNig-2 had 473 amino acids in common with other *B. subtilis* levansucrase sequences but contained two amino acid substitutions at S70P and D203N. The MENO2 sequence featured a single substitution at K174H and was missing the final lysine residue common in *B. subtilis* levansucrase proteins.

Comparative analysis between *B. subtilis* HMNig-2 and *B. subtilis* MENO2

Cell wall and capsule

Wall teichoic acids are anionic, phosphate-rich polymers related to the peptidoglycan of Gram-positive microorganisms. In the *B. subtilis* type strain 168, the predominant wall teichoic acid type is poly(glycerol phosphate) while it is poly(ribitol phosphate) in strain W23, where they are synthesized by the *tag* and *tar* gene products, respectively. Cell wall teichoic acids are essential in *B. subtilis* but could be replaceable under phosphate-limiting conditions. The putative poly ribitol phosphotransferase and ribitol-5-phosphate cytidyl transferase genes are present in *B. subtilis* HMNig-2 but absent in *B. subtilis* MENO2, while three further genes encoding components of the same subsystem, CDP-glycerol:poly

Table 1 General genomic features of *Bacillus subtilis* strains

Feature	<i>Bacillus subtilis</i> HMNig-2	<i>Bacillus subtilis</i> MENO2
Genome size (bp)	4,178,124	4,083,694
L50	1	1
GC content %	43.6	43.8
Number of subsystems	476	473
Number of coding sequences	4239	4127
Number of RNAs	129	128

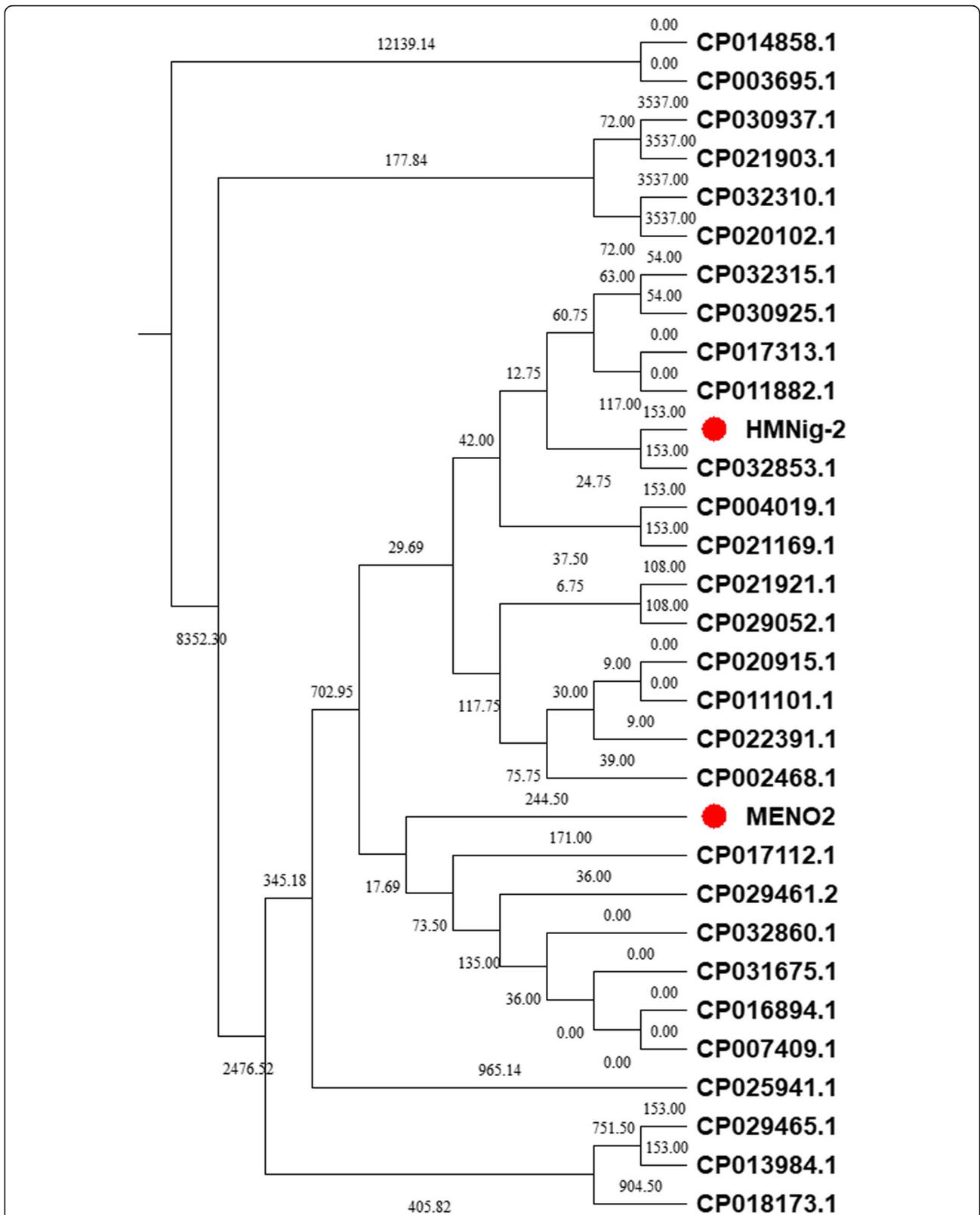
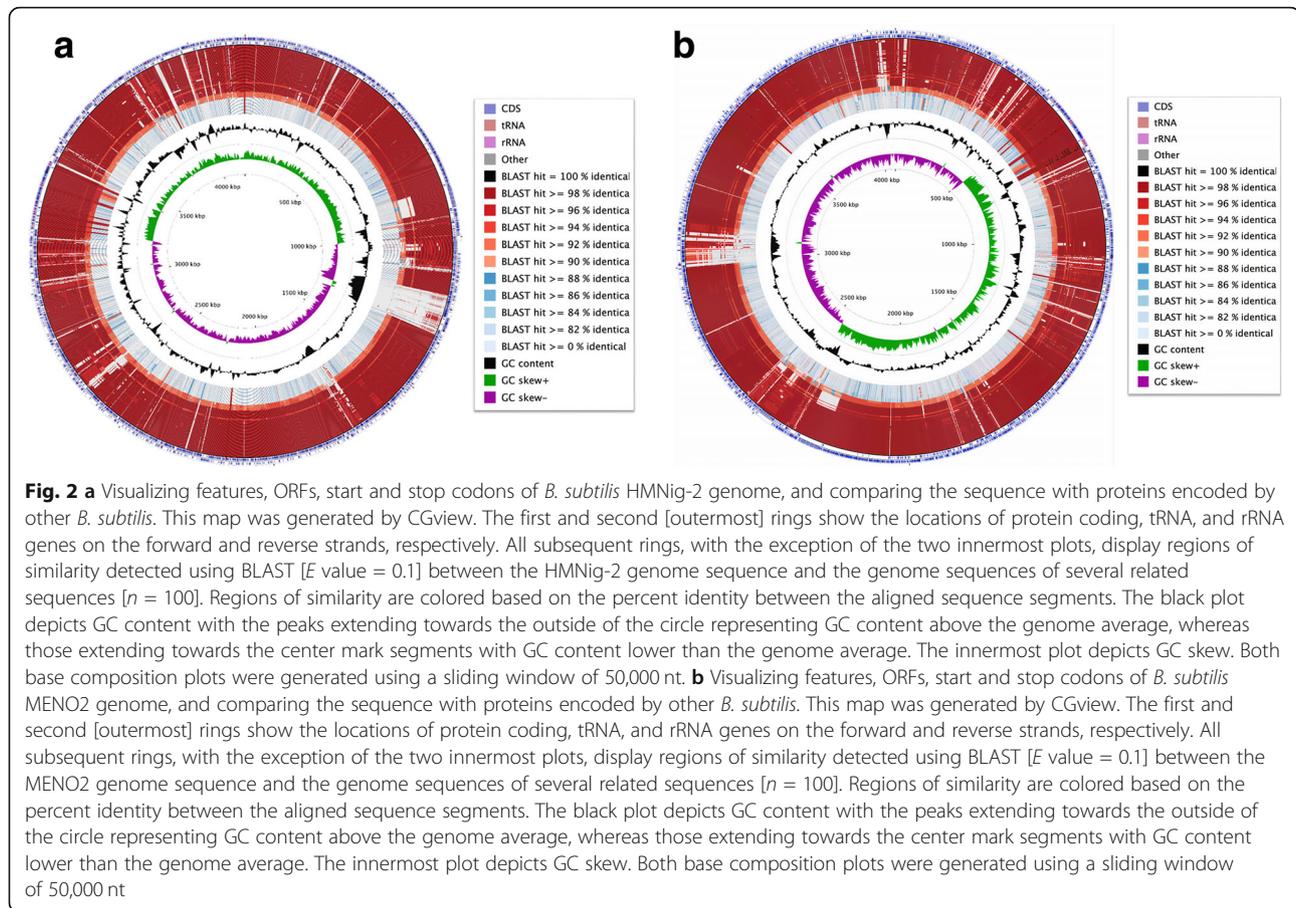


Fig. 1 Neighbor-joining phylogenetic tree of the *B. subtilis* HMNig-2 and MENO2 genome, their closest neighbors from other *B. subtilis* strains are shown. Accession numbers are given in parentheses. Phylogenetic analysis was performed using MEGA7



(glycerophosphate) glycerol phosphotransferase (EC 2.7.8.12), minor teichoic acid biosynthesis protein *GgaA*, and minor teichoic acid biosynthesis protein *GgaB*, are present in MENO2 but absent in HMNig-2.

DNA metabolism

Within this category and under the subsystem DNA repair genes, bipolar DNA helicase HerA, hypothetical protein, ATP-dependent DNA ligase, DNA-cytosine methyltransferase, very-short-patch mismatch repair endonuclease (G-T specific), and protease III precursor are present in the HMNig-2 strain but are absent in MENO2. While MENO2 strain has a prophage-associated gene responsible for recombinational DNA repair in RecT, which is absent from the HMNig-2 strain, recombinational DNA repair is closely tied to cellular replication systems, which functions to repair impairments at replication forks.

Cofactors, vitamins, prosthetic groups, and pigments

The gene encoding butyryl-CoA dehydrogenase is present within HMNig-2 strain but absent from MENO2. Butyryl-CoA dehydrogenase in *B. subtilis* is placed within the folate and pterin subcategory.

Membrane transport

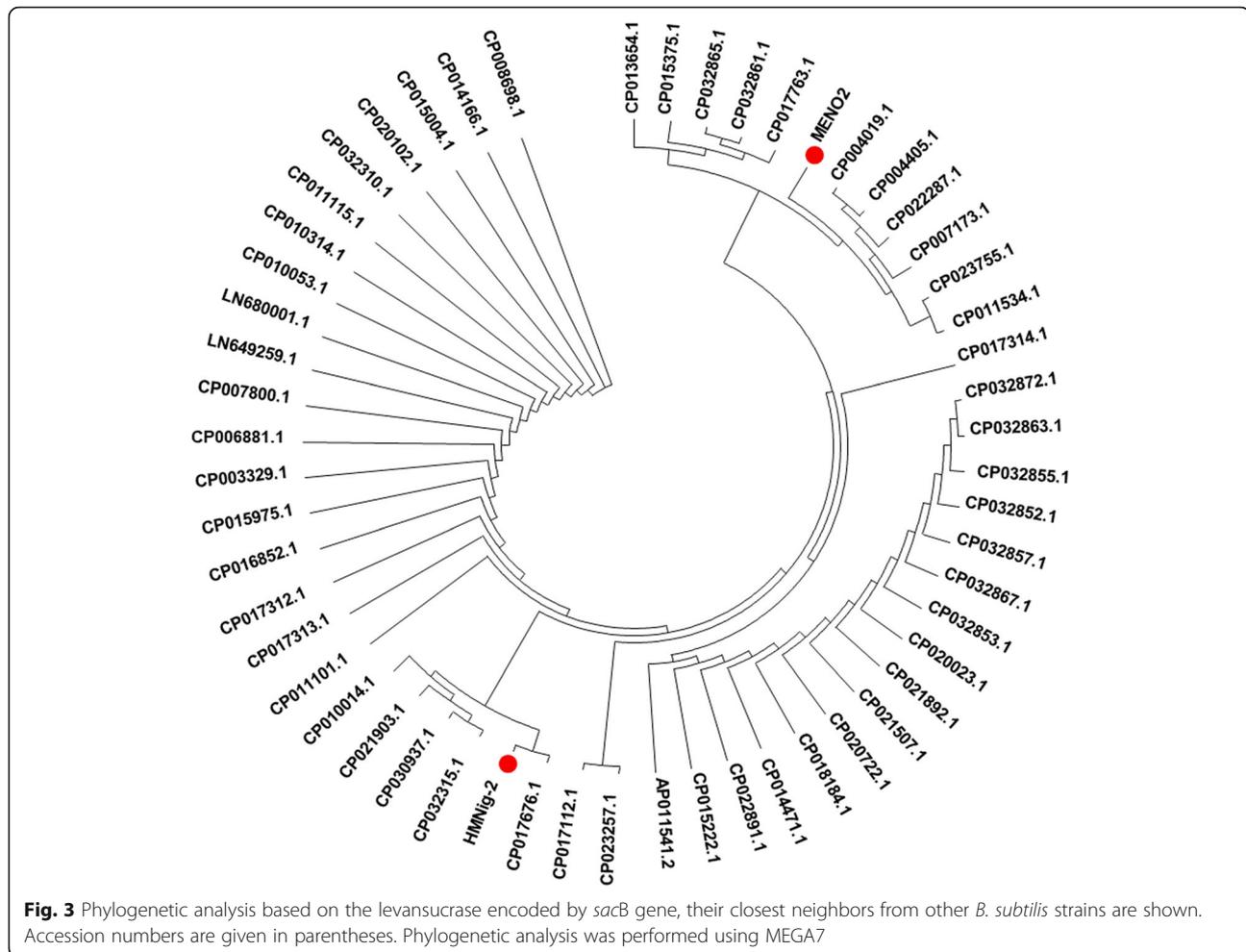
Two genes encoding CopC and CopD of the copper transport system [31] are present in strain MENO2 but are absent in HMNig-2 strain. This could be an indication of a requirement of the MENO2 strain to adapt to changes in copper availability, either when limited or to avoid toxicity.

Nitrogen metabolism

Within the subsystem for ammonia assimilation the gene encoding glutamine amidotransferase, class-II is present in MENO2 but missing from the honey HMNig-2 strain. Glutamine phosphoribosylpyrophosphate (PRPP) amidotransferase is the key regulatory enzyme of de novo purine nucleotide synthesis. Phosphoribosylamine is transformed in a series of steps to the purine nucleotide IMP, which is converted to the major end products of the pathway AMP and GMP. This enzyme is subject to tight metabolic regulation in *B. subtilis*.

Phages, prophages, transposable elements, and plasmids

Phage-associated metabolic functions encoding thioredoxin and DNA helicase were observed in the genome of HMNig-2 but absent from strain MENO2, whereas



the genes encoding the structural phage neck proteins are present from MENO2 and absent from HMNig-2 genome.

Protein metabolism

The large subunit ribosomal protein L14p (L23e) was absent from MENO2. The gene for the tRNA proofreading protein STM4549 was absent from HMNig-2. The protein processing and modification subsystem include N-linked glycosylation in bacteria, and specifically, the gene encoding a putative 4-keto-6-deoxy-*N*-acetyl-*D*-hexosaminyl-(lipid carrier) aminotransferase was absent from HMNig-2

Virulence, disease, and defense

The genes encoding for virulence factors such as hemolysin BL, non-hemolytic enterotoxin NHE, enterotoxin T, cytotoxin T, and cereulide were not present in either of the two strains. Furthermore, their genomes do not contain plasmids or the antibiotic-resistant genes against

fluoroquinolone, fosfomycin, streptogramin B, nitroimidazole, phenicol, rifampicin, sulphamide, and tetracycline.

Discussion

Bacillus spp. have been comprehensively studied due to their tough stress resistance and exceptional environmental flexibility. *B. subtilis* is a model Gram-positive bacterium that has been widely studied for research and biological purposes. It is widespread in nature and safe for humans and the environment; in addition, it has a variety of antimicrobial components and beneficial enzymes for biocontrol and pharmaceutical applications. Moreover, several strains of *B. subtilis* have been identified as probiotic candidates, and it is well known that the probiotic action is strain-specific. The *B. subtilis* HMNig-2 and MENO2 strains were previously isolated from honey and the honeybees gut microbiomes [4]. Our previous results showed their ability for high levansucrase production and levan yield. They also exhibited probiotic characteristics in vitro by expressing tolerance to gastrointestinal conditions. Furthermore, the two

strains and their levans showed effects on the immune system and protection from the *Salmonella typhimurium* infection when administrated to mouse groups, suggesting them as promising probiotic candidates [5].

Here in this work, we introduce the complete genome sequences of these valuable strains as probiotic and/or cell factory. The availability of the complete genome sequence of HMNig-2 and MENO2 not only will promote upgrade the genome database, but will also provide the opportunity to explore more genes related to different industries.

Whole-genome sequencing of various *Bacillus* strains has revealed adaptive functions in their gene contents to different environments [32–34]. In this paper, we have identified differences between our two strains HMNig-2 and MENO2, suggesting differential adaptation to the prevailing environmental conditions.

In recent years, many studies have focused on microbial sources of levansucrase (SacB), and in particular the enzyme from *B. subtilis* to reveal an important issue concerning the production of high molecular weight levan that is preferred for multiple applications [35]. It is reported that the molecular weight of the levan produced is affected by the concentration of the enzyme used in the reaction [36, 37]. The authors demonstrate that using low enzyme concentrations produces high molecular weight of levan (HMW) with an average molecular weight of 2300 kDa, whereas higher enzyme concentrations produced a low-molecular weight (LMW) levan of 7.2 kDa. *Bacillus* levansucrase supports two mechanisms dependent on the enzyme concentration, a processive reaction at low enzyme concentrations leading to HMW levan and a non-processive reaction at high enzyme concentrations that produces LMW levan [36]. Protein-product interactions are thought to be responsible for the ensuing mechanism [36], and variation in the amino acid residues responsible for these interactions will influence levan production. Therefore, strain-dependent variation in the *sacB* gene has the potential to modulate levan production.

In the other hand, with respect to their use as probiotics, they approved their safety by the absence of any virulence and antibiotic-resistant genes containing plasmid. Also, the genes encoding for virulence factors such as hemolysin BL, non-hemolytic enterotoxin NHE, enterotoxin T, cytotoxin T, and cereulide were not present in either of the two strains. Recently, the whole-genome sequencing has been used for the safety assessment of potential probiotic strains and prediction of the presence or absence of genes associated with virulence and adverse metabolite [38–40]. Therefore, in future, the complete genome sequencing would be a crucial and accurate approach for evaluating the safety and control the quality of probiotic strains.

Conclusions

In conclusion, the complete genome sequences of the prospective probiotic *B. subtilis* strains HMNig-2 and MENO2 isolated from honey and honeybee gut microbiome are reported. The strains are suitable for use as probiotics or cell factories as they do not contain any virulence or antibiotic resistance genes.

Abbreviations

B. subtilis: *Bacillus subtilis*; SacB: Levansucrase; CDS: Coding sequence; ORF: Open reading frame; kDa: Kilodaltons

Acknowledgements

This work was supported by the National Research Centre, Egypt, and Nottingham University, UK

Authors' contributions

This work was done based on two strains which were isolated during the PhD degree by NE and supervised by AH, ME, NM, MA, AA, and GA. All authors have contributed significantly, and all authors are in agreement with the content of the manuscript. NM: planned and designed the work, analyzed the genome sequence, interpreted the results, and wrote the manuscript and drafts. IC performed the genome sequence and revised and edited the manuscript. NE cultured the strains, prepared the samples, and did any technical steps. AH contributed to the presentation of the figures and worked on the manuscript. ME aided in interpreting the results and worked on the manuscript. MA aided in interpreting the results and worked on the manuscript. AA aided in interpreting the results and worked on the manuscript. GA aided in interpreting the results and worked on the manuscript. All the authors have read and approved the final manuscript.

Funding

Not applicable

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate

Not applicable

Consent for publication

All authors are in agreement with the content of the manuscript.

Competing interests

The authors declare that they have no competing of interest.

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Received: 31 March 2020 Accepted: 8 July 2020

Published online: 22 July 2020

References

1. Basualdo C, Sgroy V, Finola MS, Marioli JM (2007) Comparison of the antibacterial activity of honey from different provenance against bacteria usually isolated from skin wounds. *Vet Microbiol* 12:375–381
2. Voidarou C, Alexopoulos A, Plessas S, Karapanou A, Mantzourani I, Stavropoulou E, Fotou K, Tzora A, Skoufos I, Bezirtzoglou E (2011) Antibacterial activity of different honeys against pathogenic bacteria. *Anaerobe* 17(6):375–379

3. Tsekoura F., Alexopoulos A., Stefanis C., Papadopoulos I., Voidarou C., Bolkas S., Zoulis N., Giannakourou M., Gergopoulou A., Bezirtzoglou E. (2006) Aerobic and anaerobic bacteriology of Greek honeys. In Proceedings of the International Congress of Bioprocessing in Food Industry (ICBF) June 18-21 Patras-Greece.
4. Hamdy A.A., Elattal N.A., Amin M.A., Ali A.E., Mansour N.M., Awad G.E., Awad H.M., Esawy M.A. (2017) Possible correlation between levansucrase production and probiotic activity of *Bacillus* sp. isolated from honey and honey bee. *World J Microbiol Biotechnol* **33**(4):69. <https://doi.org/https://doi.org/10.1007/s11274-017-2231-8>.
5. Hamdy AA, Elattal NA, Amin MA, Ali AE, Mansour NM, Awad GEA, Farrag AH, Esawy MA (2018) In vivo assessment of possible probiotic properties of *Bacillus subtilis* and prebiotic properties of Levan. *Biocatal Agric Biotechnol* **13**:190–197
6. Snowdon JA, Cleaver DO (1996) Microorganisms in honey. *Int J Food Microbiol* **31**:1–26
7. Gillian M, Prest DB (1987) Microbiology of feces of the larval honey bee, *Apis mellifera*. *J Invertebr Pathol* **49**:70–75
8. Taormina PJ, Niemira BA, Beuchat LR (2001) Inhibitory activity of honey against foodborne pathogens as influenced by the presence of hydrogen peroxide and level of antioxidant power. *Int J Food Microbiol* **69**:217–225
9. Cartman ST, La Ragione RM, Woodward MJ (2007) Bacterial spore formers as probiotics for poultry. *Food Sci Tech Bull Funct Foods* **4**:21–30
10. Green DH, Wakeley PR, Page A, Barnes A, Baccigalupi L, Ricca E, Cutting SM (1999) Characterization of two *Bacillus* probiotics. *Appl Environ Microbiol* **65**:4288–4291
11. Sanders ME, Morelli L, Tompkins TA (2003) Sporeformers as human probiotics: *Bacillus*, *Sporo-lactobacillus*, and *Brevibacillus*. *Comp Rev Food Sci Food Saf (CRFSFS)* **2**:101–110
12. Urdaci MC, Bressollier P, Pinchuk I (2004) *Bacillus clausii* antimicrobial and immunomodulatory activities. *J Clin Gastroenterol* **38**(6):S86–S90
13. Spinosa MR, Braccini T, Ricca E, de Felice M, Morelli L, Pozzi G, Oggioni MR (2000) On the fate of ingested *Bacillus* spores. *Res Microbiol* **151**:361–368
14. Desai JD, Banat IM (1997) Microbial production of surfactant and their commercial potential. *Microbiol Mol Biol Rev* **61**:47–64
15. Healy MG, Devine CM, Murphy R (1996) Microbial production of biosurfactants. *Resour Conserv Recycl* **18**:41–57
16. Katz E, Demain AL (1977) The peptide antibiotics of *Bacillus*: chemistry, biogenesis, and possible functions. *Bacteriol Rev* **41**:449–474
17. Phister TG, O'Sullivan DJ, McKay LL (2004) Identification of bacilysin, chlorotetaine, and iturin A produced by *Bacillus* sp. strain CS93 isolated from pozol, a Mexican fermented maize dough. *Appl Environ Microbiol* **70**:631–634
18. Shih IL, Yu YT, Shieh CJ, Hsieh CY (2005) Selective production and characterization of Levan by *Bacillus subtilis* (Natto) Takahashi. *J Agricult Food Chem* **53**(21):8211–8215
19. Yamamoto Y, Takahashi Y, Kawano M, Iizuka M, Matsumoto T, Saeki S, Yamaguchi H (1999) In vitro digestibility and ferment ability of Levan and its hypocholesterolemic effects in rats. *J Nutr Biochem* **10**:13–18
20. Han YW, Watson MA (1992) Production of microbial Levan from sucrose, sugarcane juice and beet molasses. *J Ind Microbiol* **9**:257
21. Tajima K, Uenishi N, Fujiwara M, Erata T, Munekata M, Takai M (1997) The production of a new water soluble polysaccharide by *Acetobacter xylinum* NCI 1005 and its structural analysis by NMR spectroscopy. *Carbohydr Res* **305**:117–122
22. Tiekling M, Korakli M, Ehrmann MA, Gänzle MG, Vogel RF (2003) In situ production of exopolysaccharides during sourdough fermentation by cereal and intestinal isolates of lactic acid bacteria. *Appl Environ Microb* **69**:945–952
23. Morales-Arrieta S, Rodríguez ME, Segovia L, Lopez-Munguia A, Olvera-Carranza C (2006) Identification and functional characterization of levS, a gene encoding for a levansucrase from *Leuconostoc mesenteroides* NRRL B-512F. *Gene* **376**(1):59–67
24. Bae IY, Oh I-K, Lee S, Yoo S-H, Lee HG (2008) Rheological characterization of Levan polysaccharides from *Microbacterium laevaniformans*. *Int J Biol Macromol* **42**:10–13
25. Chiang CJ, Wang JY, Chen PT, Chao YP (2009) Enhanced Levan production using chitin-binding domain fused levansucrase immobilized on chitin beads. *Appl Microbiol Biotechnol* **82**:445–451
26. Shih IL (2010) Microbial exo-polysaccharides for biomedical applications. *Mini Rev Med Chem* **10**:1345–1355
27. Tian F, Inthanavong L, Karboune S (2011) Purification and characterization of levansucrases from *Bacillus amyloliquefaciens* in intra- and extracellular forms useful for the synthesis of Levan and fructooligosaccharides. *Biosci Biotechnol Biochem* **75**:1929–1938
28. Visnapuu T, Mardo K, Mosoarca C, Zamfir AD, Vigants A, Alamäe T (2011) Levansucrases from *Pseudomonas syringae* pv. Tomato and *P. chlororaphis* subsp. aurantiaca: substrate specificity, polymerizing properties and usage of different acceptors for fructosylation. *J Biotechnol* **155**:338–349
29. Aziz RK, Bartels D, Best AA et al (2008) The RAST server: rapid annotations using subsystems technology. *BMC Genomics* **9**:75
30. Grant J.R., Arantes A.S., Stothard P. (2012) Comparing thousands of circular genomes using the CG view comparison tool. *BMC Genomics* **13**:202. <https://doi.org/https://doi.org/10.1186/1471-2164-13-202>
31. Lawton TJ, Kenney GE, Hurley JD, Rosenzweig AC (2016) The CopC family: structural and bioinformatic insights into a diverse group of periplasmic copper binding proteins. *Biochemistry* **55**(15):2278–2290
32. Deng Y, Zhu Y, Wang P, Zhu L, Zheng J, Li R, Ruan L, Peng D, Sun M (2011) Complete genome sequence of *Bacillus subtilis* B5n5, an endophytic bacterium of *Amorphophallus konjac* with antimicrobial activity for the plant pathogen *Erwinia carotovora* subsp. *Carotovora* *J Bacteriol* **193**:2070–2071
33. Blom J, Rueckert C, Niu B, Wang Q, Borriss R (2012) The complete genome of *Bacillus amyloliquefaciens* subsp. *plantarum* CAU B946 contains a gene cluster for nonribosomal synthesis of iturin a. *J Bacteriol* **194**:1845–1846
34. He P, Hao K, Blom J, Ruckert C, Vater J, Mao Z, Wu Y, Hou M, He P, He Y, Borriss R (2012) Genome sequence of the plant growth promoting strain *Bacillus amyloliquefaciens* subsp. *plantarum* B9601-Y2 and expression of mersacidin and other secondary metabolites. *J Biotechnol* **164**:281–291
35. Porras-Domínguez JR, Rodríguez-Alegria ME, Ávila-Fernández Á, Montiel-Salgado S, López-Munguía A (2017) Levan-type fructooligosaccharides synthesis by a levansucrase-endolevanase fusion enzyme (LevB1SacB). *Carbohydr Polym* **177**:40–48
36. Raga-Carbajal E, Carrillo-Nava E, Costas M, Porras-Dominguez J, López-Munguía A, Olvera C (2016) Size product modulation by enzyme concentration reveals two distinct Levan elongation mechanisms in *Bacillus subtilis* levansucrase. *Glycobiology* **26**(4):377–385
37. Porras-Domínguez JR, Ávila-Fernández Á, Miranda-Molina A, Rodríguez-Alegria ME, Munguía AL (2015) *Bacillus subtilis* 168 levansucrase (SacB) activity affects average Levan molecular weight. *Carbohydr Polymers* **132**:338–344
38. Senan S, Prajapati JB, Joshi CG (2015) Feasibility of genome-wide screening for biosafety assessment of probiotics: a case study of lactobacillus helveticus MTCC 5463. *Probiotics Antimicro* **7**(4):249–258
39. Li B, Jin D, Evvie SE, Li N, Yan F, Zhao L, Liu F, Huo G (2017) Safety assessment of *Lactobacillus helveticus* KLD51. 8701 based on whole genome sequencing and oral toxicity studies. *Toxins* **9**(10):301
40. Li B, Zhan M, Evvie SE, Jin D, Zhao L, Chowdhury S, Sarker SK, Huo G, Liu F (2018) Evaluating the safety of potential probiotic enterococcus durans KLD56. 0930 using whole genome sequencing and oral toxicity study. *Front Microbiol* **9**:1943

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